Supplemental Figures S1-S11

Anthrax edema toxin disrupts distinct steps in Rab11-dependent junctional transport. Guichard A, Jain P, Moayeri M, Schwarz R, Chin S, Zhu L, Cruz-Moreno B, Liu JZ, Aguilar B, Hollands A, Leppla SH, Nizet V, Bier E. PLoS Pathog 2017; 13:e1006603.



Figure S1. Activated Rab11 (Rab11*) preferentially accumulates at AJs in Drosophila wing imaginal discs. (A-C) Expression of Rab11wt in 1096GAL4>Rab11wtYFP wing discs. (A) Rab11wtYFP detected with a rabbit anti-GFP antibody appears as a peppered stain near the apical surface. (B) D-Ecad/GFP double stain. (C) corresponding D-Ecad stain marking AJs. (D-F) Expression of Rab11* in 1096GAL4>Rab11*YFP wing discs. (D) Rab11*YFP detected with a rabbit anti-GFP antibody. In addition to a peppered apical stain, Rab11* shows a distinctive net-like pattern at cell borders. (E) D-Ecad/GFP double stain, revealing that Rab11*YFP tends to accumulate at the AJs. (F) Corresponding D-Ecad single stain.



Figure S1. RNAi transgenes specifically block expression of cognate proteins Sec15, Crag, and Arf6. (A) A 1096GAL4>UAS-Sec15GFP salivary gland, stained with an anti-GFP antibody. (B) Sec15GFP expression in strongly inhibited by co-expression of a Sec15RNAi construct. (C) A 1096GAL4>UAS-CragHA salivary gland stained with an anti-HA antibody. (D) CragHA expression is suppressed by co-expression of a CragRNAi construct. (E) A 1096GAL4>UAS-Arf6Myc gland stained with an anti-Myc antibody. (F) Arf6Myc expression is blocked by co-expression of an Arf6RNAi construct.



Figure S3. Blocking expression of Sec15, but not of the Rab11GEF Crag, prevents Rab11*YFP targeting to cell junctions in Drosophila salivary glands. (A-C) Rab11*YFP detected with a rabbit anti-GFP antibody in salivary glands. (A) Rab11*YFP selectively accumulates at the AJs in 1096GAL4>Rab11*YFP salivary glands. (B) Rab11* distribution is unchanged in 1096GAL4>Rab11*YFP+CragRNAi glands. (C) Rab11*YFP fails to accumulate at the AJs in 1096GAL4>Rab11*YFP +Sec15RNAi salivary glands.



Figure S4. EF prevents Rab11* accumulation at AJs. Images from experiment described in Fig 1, panels E, F, H and I, were analyzed to quantify the effect of EF on junctional accumulation of Rab11*. Individual image crops from intercellular boundaries were generated. For each crop, average fluorescence was determined in ImageJ, and normalized to the average fluorescence found inside the corresponding cell. EF expression significantly reduces Rab11* accumulation at intercellular borders, (p<0.0001).



Figure S5. Inhibition of Rab11 function in salivary glands leads to abnormal accumulation of D-Ecad around AJs, and intercellular gaps. (A-D) Salivary glands stained with an anti-D-Ecad antibody. (A) A wild-type salivary gland showing D-Ecad accumulation at AJs. (B) A SglGAL4>Rab11DN salivary gland, in which Rab11 inhibition in this tissue leads to D-Ecad accumulation in broad zones around intercellular gaps. (C-D) Higher magnifications. (C) A wildtype salivary gland showing D-Ecad forming AJs (arrows). (D) A SglGAL4>Rab11DN salivary gland, revealing gaps between cells, and broad accumulation of D-Ecad around them (arrows). D-Ecad fails to form AJs.



Figure S6. Reduction of Epac -but not PKA- levels, suppresses the EF wing phenotype. (A-F) wings of the following genotypes: (A) Wild-type (+/+). (B) 1096GAL4>EF. (C) 1096GAL4>EF+EpacRNAi. Inhibition 1096GAL4>EpacRNAi. (D) of Epac expression potentlyreduces the EF phenotype. (E) PKA-C1B10/+ (B10 is a loss -of-function allele of PKA). (F) 1096GAL4>EF; PKA-C1B10/+. Reduction of PKA-C1 levels, either in a heterozygote loss-offunction PKA-C1 alleles (B10/+) or in flies expressing a dominant negative form of PKA-C (C1-DN), does not obviously alter the EF phenotype. (G) The surface areas of wings of the indicated genotypes were measured in Photoshop. Results were plotted as a histogram, with relevant pvalues indicated. EF expression reduces wing size significantly compared to widl-type (wt) (****p<0.0001). EpacRNAi ameliorates the EF phenotype (****p<0.0001).



Figure S7. EF does not disrupt dRip11DN/Rab11 co-localization in salivary glands. (A-C) 1096GAL4>Rip11DN-GFP salivary glands, stained with (A) a rabbit anti GFP antibody, (B) a mouse anti Rab11 antibody, and (C) both antibodies, showing that Rab11 and Rip11DN-GFP co-localize in punctate vesicles. (D-F) 1096GAL4>Rip11DN+EF salivary glands stained with a rabbit anti-GFP antibody (D), a mouse anti-Rab11 antibody (E), and both antibodies (F), showing that Rab11 and Rip11DN still co-localize in EF-expressing glands. However, EF alters the distribution of both proteins, transforming small punctate staining into a ring-shaped halo surrounding secretory vesicles.



Figure S8. ET treatment reduces Rab11/Rip11 co-localization in MDCK cells. Co-localization

between Rip11-GFP and DsRed-Rab11A in co-transfected MDCK cells measured by the Pearson's correlation coefficient (PCC) is reduced by ET treatment (n = 43, p<4.85X10-9).



Figure S9. ET treatment reduces Sec15/Rab11* and Rab11*/Rip11 co-localization in HBMEC cells. (A-C) HBMECs, untreated. (D-F) HBMECs treated with ET for 6hours. Co-localization of Rab11* with Sec15 (B and E) and Rab11* with Rip11 (C and F) can be visualized following transfection of cells with Sec15-GFP. High-level expression of Sec15-GFP, and staining with an anti-Rab11* antibody (A) reveals a high degree of Sec15/Rab11* co-localization (B). In ET-treated cells, this co-localization is severely reduced (E). A double label Rab11*/Rip11 stain, reveals Rab11*/Rip11 co-localization (C), which is also abrogated by ET (F).



D-Ecad stain intensity at apical junctions

Figure S10. Arf6RNAi rescues normal apical D-Ecad levels in EF-expressing wing discs. Apical levels of D-Ecad in wing discs was measured using ImageJ. Arf6RNAi restores normal levels of apical D-Ecad in 1096GAL4>EF+Arf6RNAi discs (p<0.0001). Arf6RNAi does not notably affect apical levels of D-ECad. Surface areas of wings of the same genotypes were also measured, and Arf6RNAi showed a modest yet significant restorative effect in EF-expressing wings (in 1096GAL4>EF+Arf6RNAi wings, p<0.05).



Figure S11. ET treatment reduces the levels of cadherins and Rab11 in HBMECs. (A-B) Western blot analysis of HBMECs cells. (A) Rab11A (~28 kD) and pan-Cadherin (~97 kD) levels are severely decreased in ET-treated cells (24hrs), while control actin (~42 kD) levels are only slightly reduced. (B) dcAMP, a cell-permeant stable analog of cAMP that is insensitive to phosphodiesterase, also causes a severe loss of Rab11, suggesting that the reduction of Rab11 levels induced by ET is mediated by cAMP.