

## Supplementary Information

### Observing the Invisible through Imaging Mass Spectrometry, a Window into the Metabolic Exchange Capacity of Microbes

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**Supplementary Table 1 Growth conditions and MALDI signals of the strains used in this study.** Column definitions: (i) Strains. Indicates the specific name of the organism used in the study. (ii) Medium. Indicates the specific medium used to grow the organism. (iii) Growth time. Indicates the growth time in days (d) of the organism before analysis. (iv) MALDI-I mass range. Indicates the mass-to-charge (m/z) range monitored during data collection. (v) Mass-to-charge signals observed. Ions with spatial distributions either colony bound or released into the agar and had distinct patterns to the matrix peaks were recorded as ion candidates. (vi) Identified ions. Indicates ions that were structurally verified by tandem mass spectrometry including parent masses [M] and salt adducts [Na or K]. (vii) References. List of publications with information either on the specific strain used or the resolved molecules.

Strains	Medium	Growth time	MALDI-I (m/z range)	m/z signals observed	The ions identified (SI-4) annotated m/z signals	References
<i>Beauveria bassiana</i> ATCC 7159	ISP-2	6d	(200-3700)	204, 217, 222, 231, 236, 242, 246, 258, 260, 274, 281, 370, 296, 397, 412, 428, 461, 477, 520, 522, 575, 577, 579, 591, 593, 597, 600, 613, 615, 617, 629, 631, 654, 656, 804, 807, 823, 932, 948	Beauvericin ([M+Na] <sup>+</sup> , 807; [M+K] <sup>+</sup> , 823), Bassianolide ([M+Na] <sup>+</sup> , 932; [M+K] <sup>+</sup> , 948)	Xu, 2009
	PDA	6d	(200-3400)	231, 393, 397, 412, 520, 577, 600, 656, 688, 767, 789, 807, 823, 932, 948		
<i>Chaetomium chiversii</i> CS-36-62	ISP-2	5d	(200-1600)	404, 409, 490, 492, 505, 507, 530, 542, 546, 557, 562, 611, 633, 649, 667, 689, 705		Wang, 2009
	PDA	5d	(200-1600)	404, 409, 412, 486, 520, 542, 546, 404, 409, 490, 492, 505, 507, 530, 542, 546, 557, 562, 611, 633, 649, 667, 689, 705, 611, 633, 649, 667, 689, 705		
<i>Fusarium verticillioides</i> 5777	ISP-2	5d	(300-3600)	664, 1311, 1327, 655, 677, 693		Zhu, 2008
	PDA	5d	(300-3600)	751, 760, 766, 893 1311, 1327, 655, 677, 693		
<i>Fusarium</i> sp. CNL 292	ISP-2	5d	(350-3000)	412, 645, 647, 663, 673, 675, 691, 802, 609, 625		Belofsky, 1999
	A1	5d	(350-1600)	412, 496, 562, 581, 595, 587, 609, 625		
<i>Acremonium</i> sp. CNC890	ISP-2	5d	(300-3600)	515, 520, 531, 764, 780, 796		This study
	A1	5d	(300-3600)	476, 750, 764, 766, 780, 796, 876		

<i>Fusarium</i> sp.CNC 477	ISP-2	5d	(200-3600)	258, 296, 493, 515, 531, 547		Renner 1998
	A1	7d	(350-1600)	493, 515, 531, 547, 747		
Thermofungus C030813_30	PDA		(350-1600)	460, 476, 546,688, 739, 767, 842, 863, 881		This study
Thermofungus C030813_40	PDA		(350-1600)	445, 460, 589, 656, 672, 751, 767, 771, 786, 881		
<i>Salinispora arenicola</i> CNS205	A1	15d	(350-1600)	354, 365, 447, 458, 497, 527, 540, 556, 655, 672, 729, 751, 767, 940		Schultz 2008
<i>Salinispora pacifica</i> CNS 143	A1	11d	(250-1600)	None		Jensen 2006
<i>Salinispora pacifica</i> CNT 133	A1	11d	(250-1600)	495		This study
<i>Salinispora arenicola</i> CNH 643	A1	11d	(250-1600)	457, 467, 489, 556		This study
<i>Salinispora arenicola</i> CNT 088	A1	11d	(250-1600)	354, 557		Asolkar 2009
Actinomycete CNS 575	ISP-2	16d	(250-1600)	879, 901, 917	Etamycin of [M+H] <sup>+</sup> , 879; [M+Na] <sup>+</sup> , 901; and [M+K] <sup>+</sup> , 917.	Haste 2010
	A1	11d	(250-1600)	623, 901, 917		
<i>Streptomyces</i> sp. SPB74	ISP-2	10d	(300-3600)	637, 682, 857, 877, 909, 1044, 1046, 1105		Oh 2009
<i>Streptomyces</i> sp. AA#4	ISP-2	7d	(300-3600)	655, 666, 682, 693, 720, 777, 877, 1059, 1073, 1089, 1102, 1111, 1182, 1354.		This study
<i>Streptomyces</i> sp. Mg1	ISP-2	8d	(300-3600)	668, 973, 1007, 1988, 1904, 1969, 1985, 2003		This study
<i>Streptomyces</i> sp. WASP	ISP-2	7d	(300-3600)	582, 618, 634, 656, 672, 705		This study
<i>S.coelicolor</i> A3(2)	ISP-2	8d	(300-3600)	424, 392, 394, 1536, 2027, 2065	Prodiginines(392, 394), CDA(1536), sapB(2027, 2065)	Yang 2009
<i>S. roseosporus</i> NRRL15998	ISP-2	8d	(300-3600)	799, 815, 831, 845, 848, 862, 864, 878, 893, 972, 2237, 2253		Debono 1987
<i>S. hygroscopicus</i> ATCC53653	ISP-2	10d	(300-3600)	1038, 1060, 1076, 1102, 1118, 1136, 1462, 1476, 1490, 1548, 1562, 1573, 1587, 1601, 1615, 1629, 1639, 1653, 1667		Lam 1990
<i>S. albus</i> J1074	ISP-2	9d	(300-3600)	486, 655, 732, 950, 1061, 1077, 1093, 1109, 2266, 2282, 2304, 1320		Lombó 2006
<i>S. ghanaensis</i> 14672	ISP-2	7d	(300-3600)	666, 739, 809, 823, 841, 2014, 2052		Endler 1998

<i>S. clavuligerus</i> ATCC 53653	ISP-2	9d	(300-3600)	2181, 2197, 2219, 2235		
<i>S. roseosporus</i> NRRL11379	ISP-2	11d	(300-3600)	1207, 1209, 1221, 1223, 1237, 1673, 1687, 1701, 1705, 1719, 1733, 1747, 1749, 2236, 2252		Huber 2005
<i>S. pristinispiralis</i> ATCC25486	ISP-2	8d	(300-3600)	641, 645, 650, 673, 711, 2014, 1036, 2052, 2074, 2090		This study
<i>S. viridochromogenes</i> 40736	ISP-2	7d	(300-3600)	992, 1689, 1711, 1727, 2068, 2169		Blodgett 2005
<i>Streptomyces sviveus</i> 29083	ISP-2	7d	(300-3600)	1698, 1812, 1826, 1954, 1870, 1888, 1910, 1926, 1948, 1968, 1984, 2084, 2106, 2122, 2138, 2154		This study
<i>Streptomyces sp.</i> SPB 78	ISP-2	7d	(300-3600)	509, 540, 553, 587, 589, 2251, 2267, 2289, 2305		This study
<i>Kutzneria sp.</i> 744	ISP-2	7d	(300-3600)	867, 883, 888, 905, 907, 921, 923, 942, 944, 958, 960, 983, 998, 1065, 1073, 1087, 1089, 1103, 1131, 1133, 666, 682, 684, 893, 1189		Broberg 2006
<i>Actinoplanes teichomyceticus</i> PSK0532	ISP-2	7d	(300-3600)	666, 682, 684, 893, 1189		This study
<i>Streptoverticillium griseoverticillatum</i> ATCC 31499	ISP-2	8d	(300-3600)	693, 709, 809, 1728, 1750, 1766, 2025, 2041, 2057, 2063, 2079, 2101, 2117, 2139, 2155	Cinnamycin ([M+H] <sup>+</sup> , 2041), [M+Na] <sup>+</sup> , 2063; [M+K] <sup>+</sup> , 2079)	Kaletta 1991
<i>Pseudomonas aeruginosa</i> PAO	ISP-2	2d	(500-2000)	577, 591, 616, 625, 651, 740, 746, 756, 760, 770, 778, 794, 798, 808, 830, 714, 1334	Polyglutamate (714),	Meyer 1996
<i>Staphylococcus aureus</i> USA300	ISP-2	2d	(300-3600)	537, 551, 565, 594, 748, 762, 775, 785, 797, 799, 804, 814, 841, 886, 904, 916, 931, 941, 946, 957, 970, 974, 775, 871, 1013, 1338, 1500	δ-toxin (3006)	Schlievert 2010
<i>Nostoc sp.</i> PCC 7120	BG-11	12d	(300-3600)	775, 871, 1013, 1338, 1500	Pheophytin A ([M+H] <sup>+</sup> , 871)	Dembitsky 2005
<i>Lysobacter enzymogenes</i> C3	ISP-2	4d	(300-3600)	571, 573, 587, 589, 1106, 1108, 1275, 1277, 1484, 1500, 1516, 1536, 1549, 1563, 1571, 1577, 1581, 1585, 1601, 1615, 1633, 1649, 1715, 1729, 1743, 1757, 1771, 1785, 1799, 1813, 511, 533, 549, 513, 535, 551	Maltophilin ([M+H] <sup>+</sup> , 511), [M+Na] <sup>+</sup> , 533; [M+K] <sup>+</sup> , 549). Dihydromaltophilin ([M+H] <sup>+</sup> , 513), [M+Na] <sup>+</sup> , 535; [M+K] <sup>+</sup> , 551)	Yu 2007
<i>Bacillus subtilis</i> 3610	ISP-2	2d	(300-3600)	655, 714, 1031, 1045, 1047, 1059, 1061, 1075, 1517, 1531, 1545	Polyglutamate (714), Surfactin(1075), Plipastatin (1545)	Yang 2009
<i>Bacillus pumilus</i> CNJ762	ISP-2	3d	(200-3800)	551, 616, 655, 844, 1051, 1059, 1073, 1087, 714, 1075, 1089, 1103, 1117	Polyglutamate (714), Surfactin (1075)	This study
	A1	5d	(300-3600)	1059, 1073, 1081, 1087, 1095, 1101, 1123, 1133, 1089, 1103, 1117		

<i>Rhizobium leguminosarum</i> S36	ISP-2	6d	(300-3600)	415, 563, 814, 843	This study
<i>Pseudomonas fluorescens</i> PFO-1	ISP-2	3d	(300-3600)	395, 428, 563, 575, 578, 585, 590, 592, 604, 616, 626, 648, 652, 664, 678, 680, 700, 715, 733, 743, 747, 761	This study
<i>Mycobacterium smegmatis</i> MC2 155 ISP-2	ISP-2	6d	(300-3600)	715, 1275, 1287, 1301	Billman-Jacobe 2002

### References to accompany Supplementary Table 1

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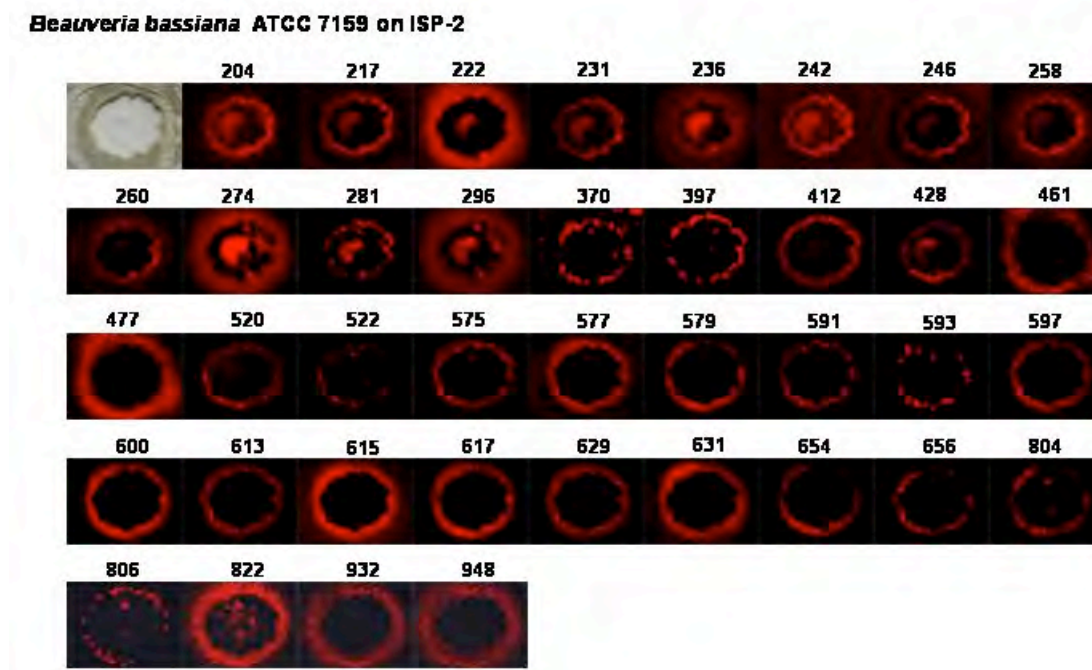
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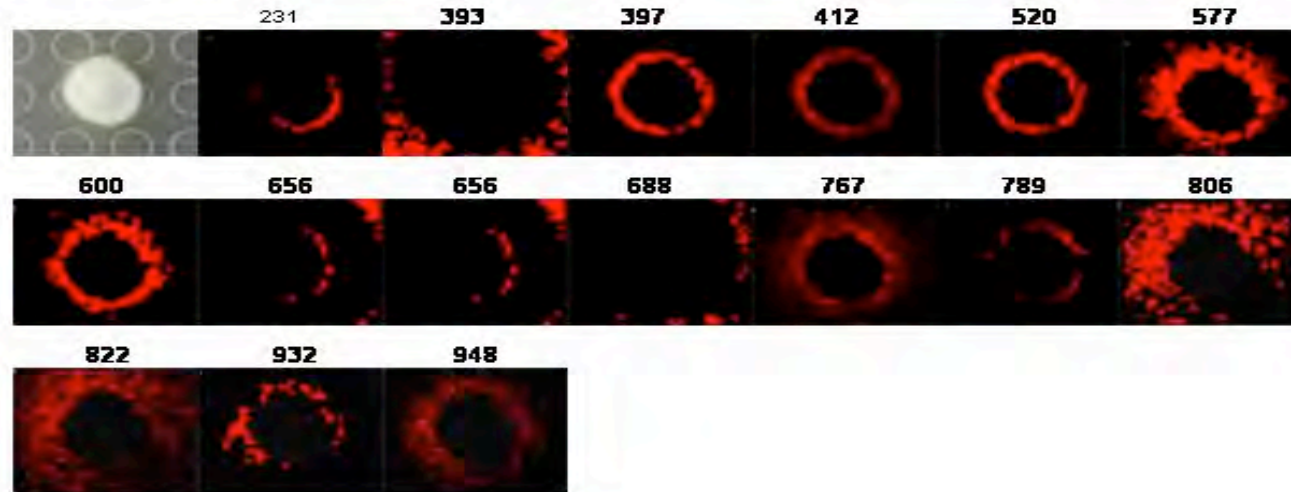


Supplementary Figure 1. The IMS images of strains indicated in Supplementary Table 1.



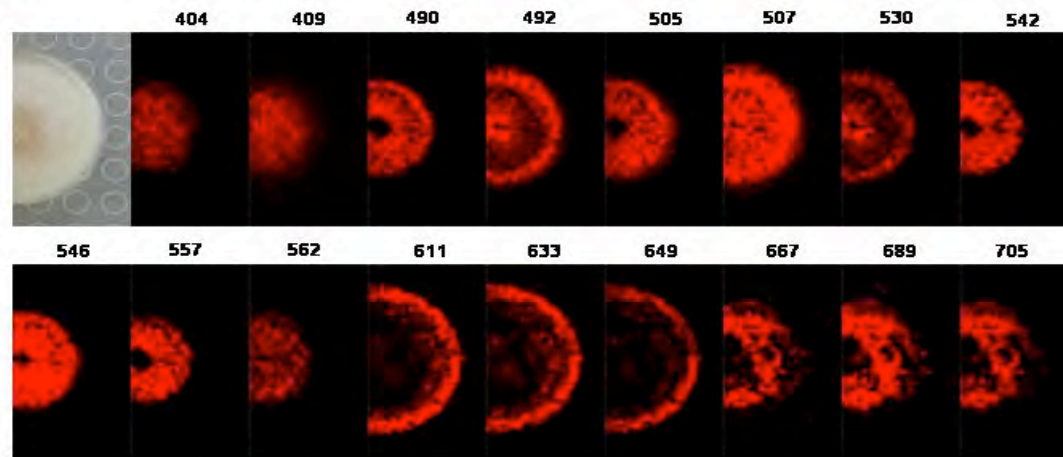
IMS 1. *Beauveria bassiana* grown on ISP-2 media. The ion at 806 Da was identified to be the [M+Na] of Beauvericin by tandem mass spectrometry (Supplemental Figure 2). IMS did not detect the [M+H] ion corresponding to Beauvericin. Liquid-liquid extraction of the organism showed the presence of the [M+H] that was resolved at 3.8 ppm mass error by FTICR-MS. Liquid-liquid extraction showed the presence of the [M+K] at 7.8 ppm mass error. The ion at 932 Da was identified to be the [M+Na] of Bassianolide by tandem mass spectrometry (Supplemental Figure 2).

***Beauveria bassiana* ATCC 7159 on PDA**



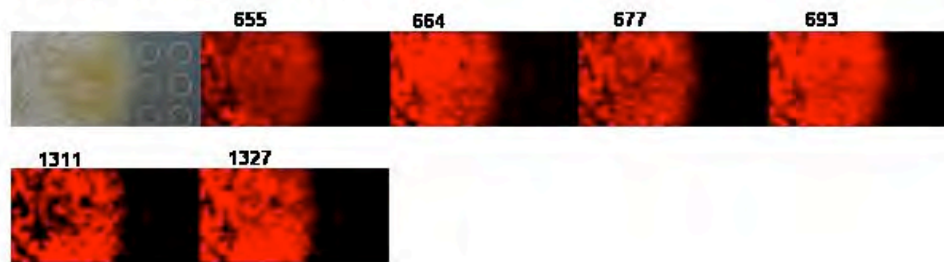
IMS 2. *Beauveria bassiana* grown on ISP-2 media. The ion at 806 Da was identified to be the [M+Na] of Beauvericin by tandem mass spectrometry (Supplemental Figure 2). IMS did not detect the [M+H] ion corresponding to Beauvericin. Liquid-liquid extraction of the organism showed the presence of the [M+H] that was resolved at 3.8 ppm mass error by FTICR-MS. Additionally, the liquid-liquid extraction showed the presence of the [M+K] at 7.8 ppm mass error. The ion at 932 Da was identified to be the [M+Na] of Bassianolide by tandem mass spectrometry (Supplemental Figure 2).

***Chaetomium chiversii* CS-36-62 on ISP-2**

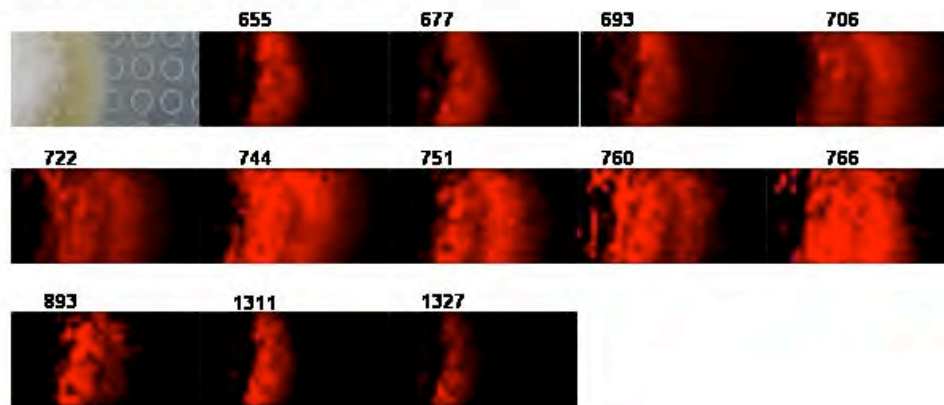


IMS 3. *Chaetomium chiversii* grown on ISP-2 media.

***Fusarium verticillioides* 5777 on ISP2**

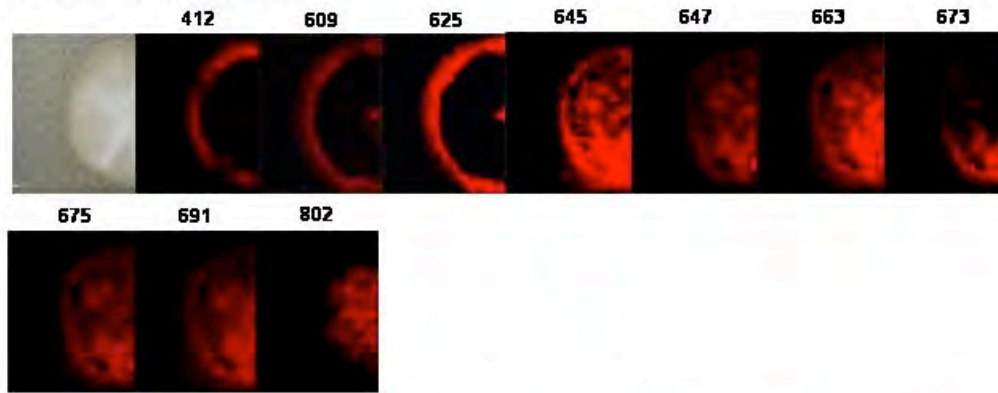


***Fusarium verticillioides* 5777 on PDA**

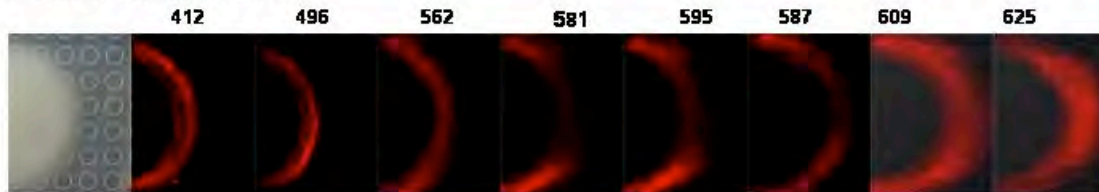


IMS 4. *Fusarium verticillioides* grown on ISP-2 media (top) and PDA media (bottom).

***Fusarium* sp. CNL292 on ISP-2**

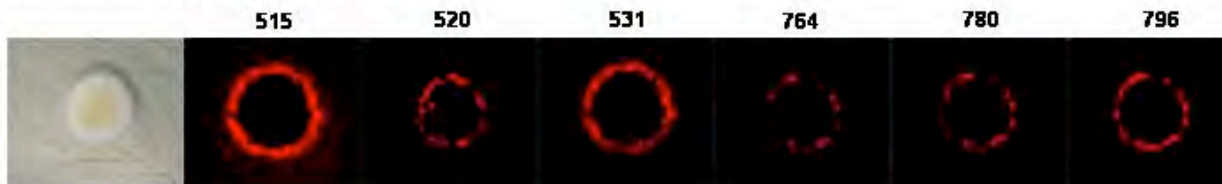


***Fusarium* sp. CNL292 on A1**

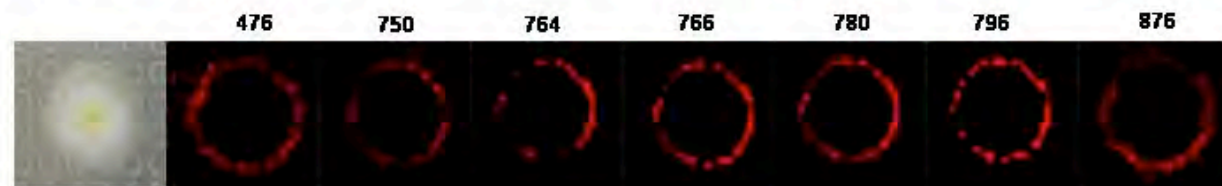


IMS 5. *Fusarium* sp. CNL292 grown on ISP-2 media (top) and A1 media (bottom).

***Acremonium* sp. CNC890 on ISP-2**



***Acremonium* sp. CNC890 on A1**

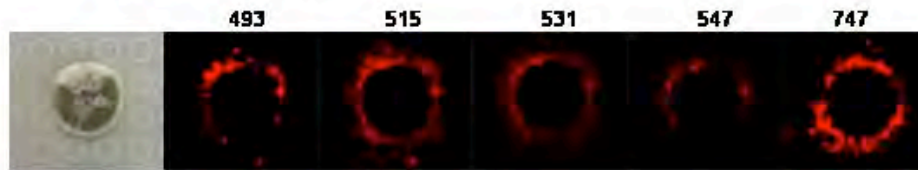


IMS 6. *Acremonium* sp. CNC890 grown on ISP-2 media (top) and A1 media (bottom).

***Fusarium* sp.CNC477 on ISP-2**

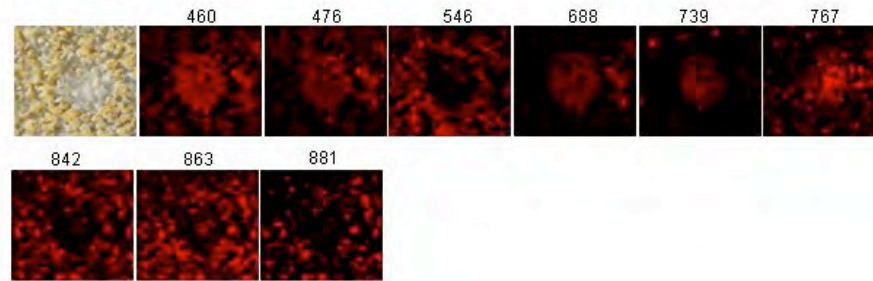


***Fusarium* sp.CNC477 on A1**

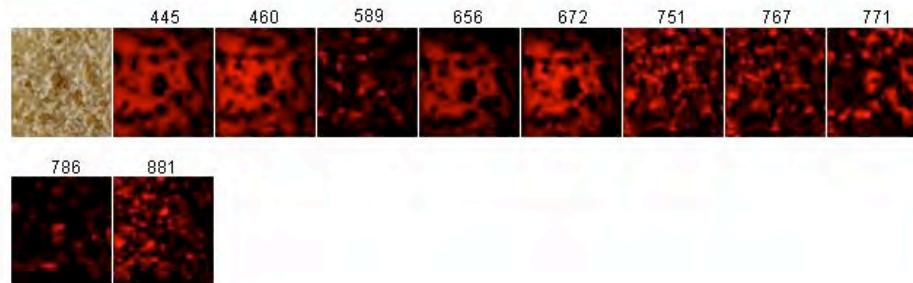


IMS 7. *Fusarium* sp. CNC477 grown on ISP-2 media (top) and A1 media (bottom).

**Thermo-fungus C030813-30**



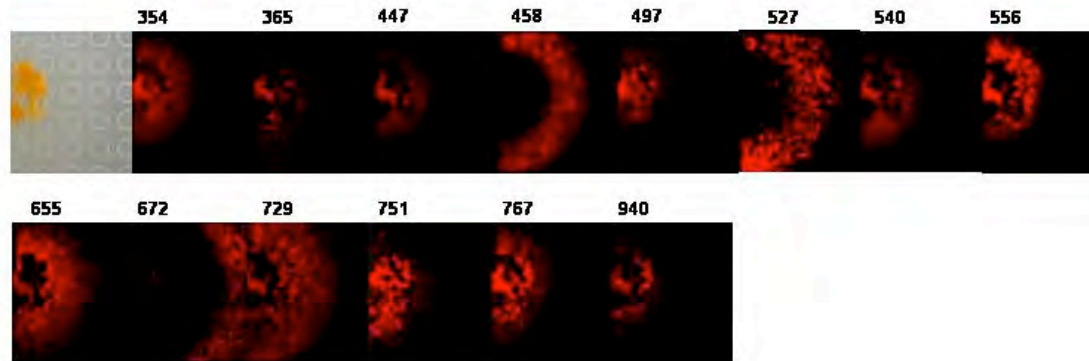
**Thermo-fungus C030813-34**



IMS 8. Thermo-fungus grown on PDA media.

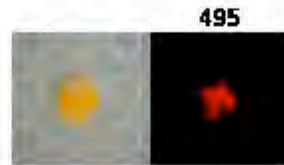


***Salinispora arenicola* CNS205 on A1**



IMS 9. *Salinispora arenicola* CNS205 grown on A1 media.

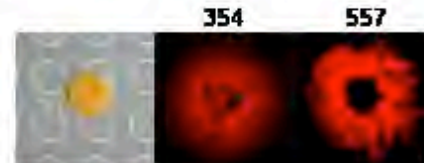
***Salinispora pacifica* CNT 133 on A1**



***Salinispora arenicola* CNH 643 on A1**

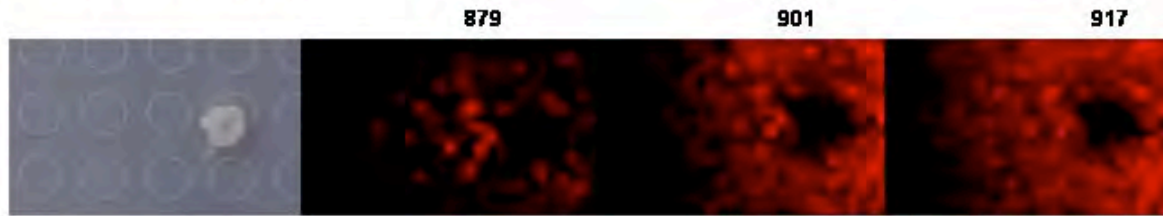


***Salinispora arenicola* CNT 088 on A1**



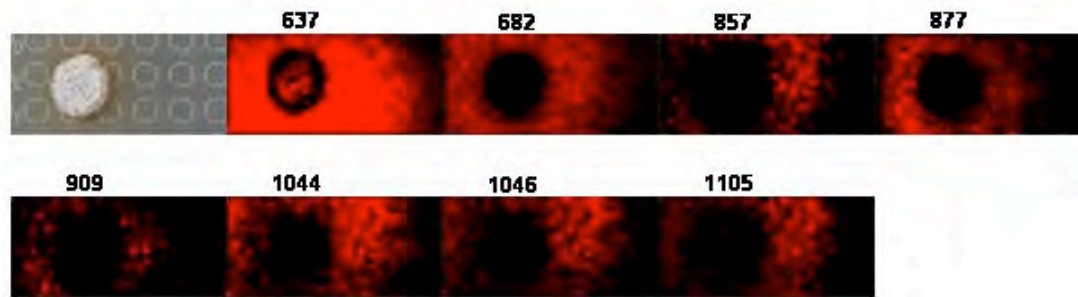
IMS 10. *Salinispora pacifica* CNT 133 grown on A1 media (top). *Salinispora arenicola* CNH 643 grown on A1 media (middle). *Salinispora arenicola* CNT 088 grown on A1 media.

**Actinomycete CNS 575 on ISP-2**



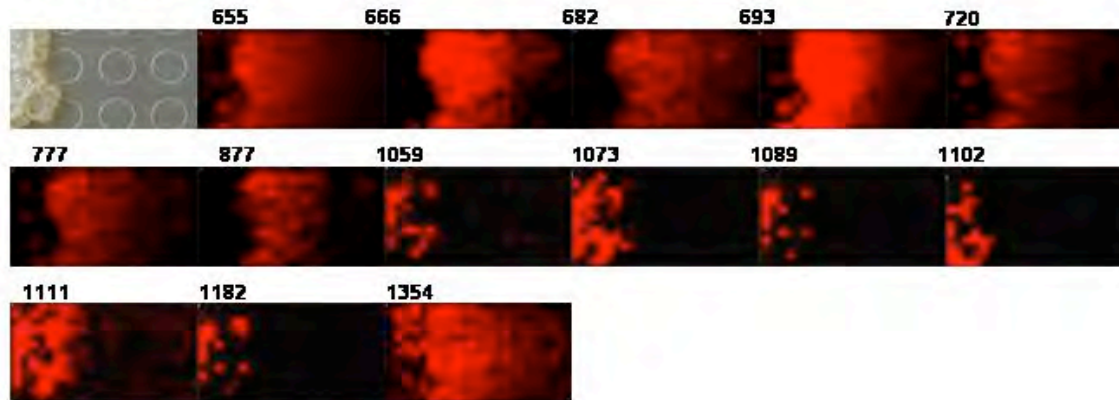
IMS 11. Actinomycete CNS 575 grown on ISP-2 media. The ion at 879 Da was identified to be the [M+H] of Etamycin by tandem mass spectrometry and FTICR-MS analysis of the intact mass (Supplemental Figure 2).

**Streptomyces sp. SPB74 on ISP-2**



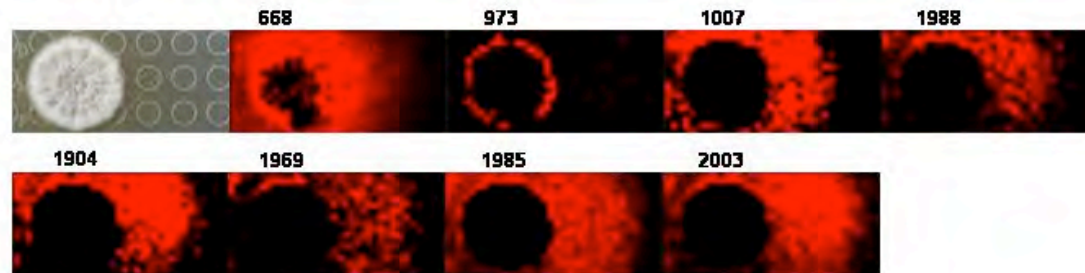
IMS 12. Streptomyces sp. SPB74 grown on ISP-2 media.

**Streptomyces sp. AA#4 on ISP-2**



IMS 13. Streptomyces sp. AA#4 grown on ISP-2 media.

**Streptomyces sp. Mg1 on ISP-2**



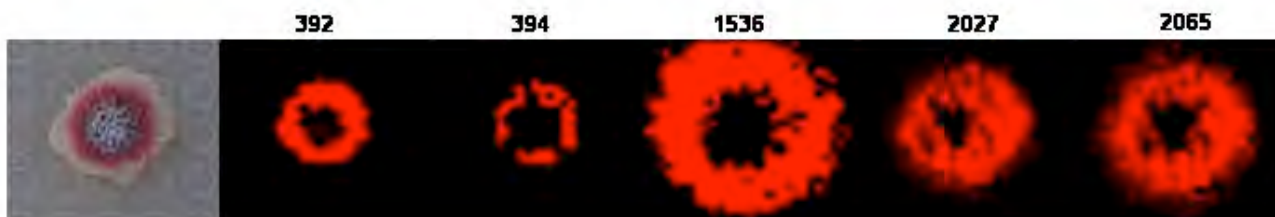
IMS 14. Streptomyces sp. Mg1 grown on ISP-2 media.

**Streptomyces sp. WASP on ISP-2**



IMS 15. Streptomyces sp. WASP grown on ISP-2 media.

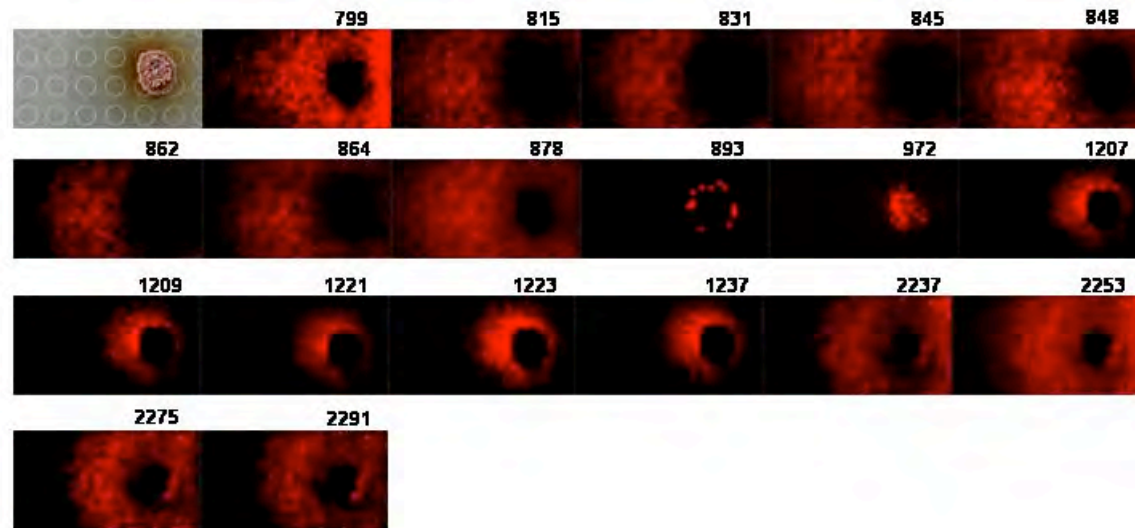
**Streptomyces A3(2) on ISP-2**



IMS 16. *Streptomyces coelicolor* grown on ISP-2 media. The ion at 392 Da was identified to be the [M+H] of Streptorubin B by tandem mass spectrometry (Supplemental Figure 2). The ion at 1536 Da was identified to be the [M+K] of CDA (Supplemental Figure 2). IMS did not detect the [M+H] ion corresponding to CDA. Liquid-liquid extraction of the organism showed the presence of the [M+H] CDA ion that was resolved at 17.5 ppm mass error by MALDI-MS. Additionally, the liquid-liquid extraction showed the presence of the [M+K] at 6.5 ppm mass error.

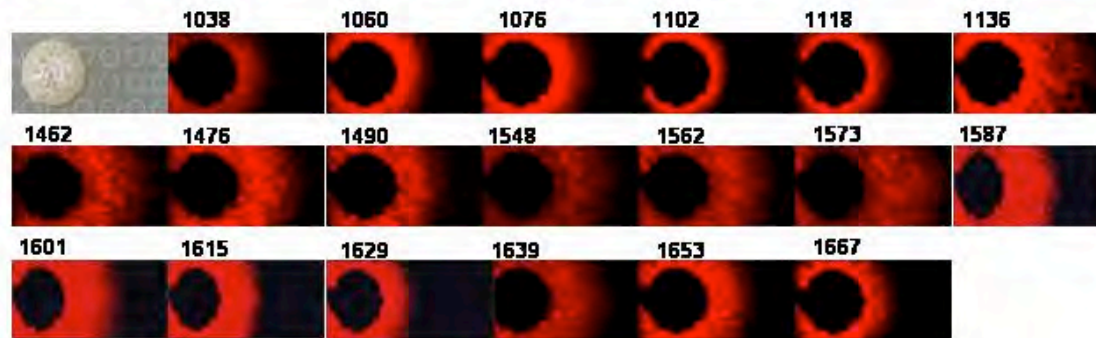


***Streptomyces roseosporus* NRRL15998 on ISP-2**



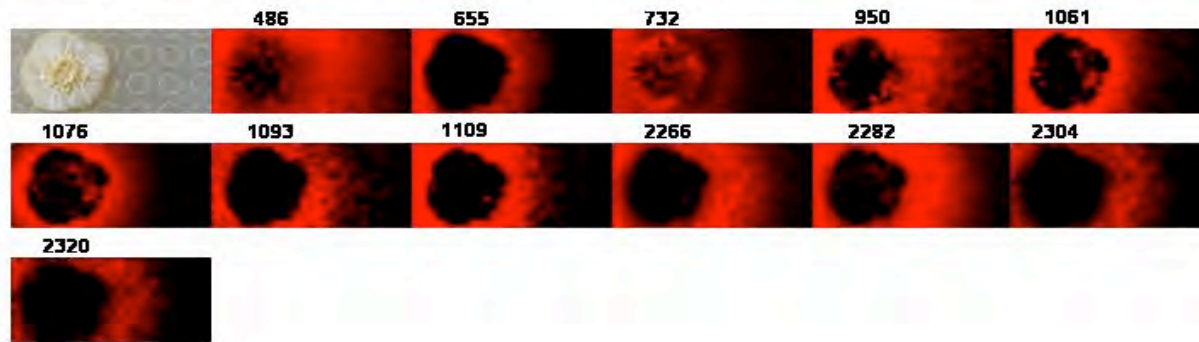
IMS 17. *Streptomyces roseosporus* grown on ISP-2 media.

***Streptomyces hygroscopicus* ATCC53653 on ISP-2**



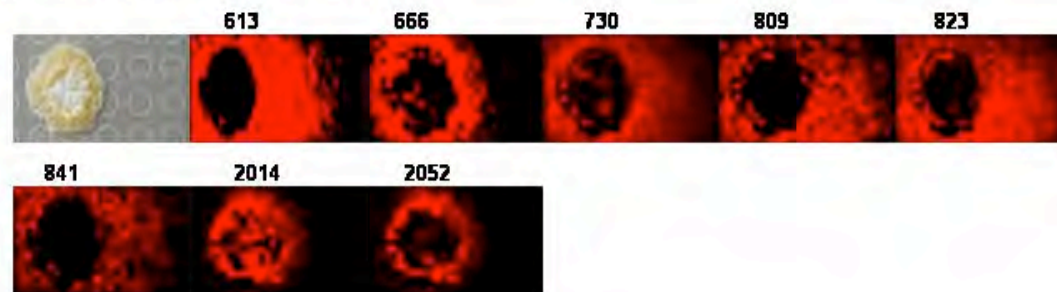
IMS 18. *Streptomyces hygroscopicus* grown on ISP-2 media.

***Streptomyces albus* J1074 on ISP-2**



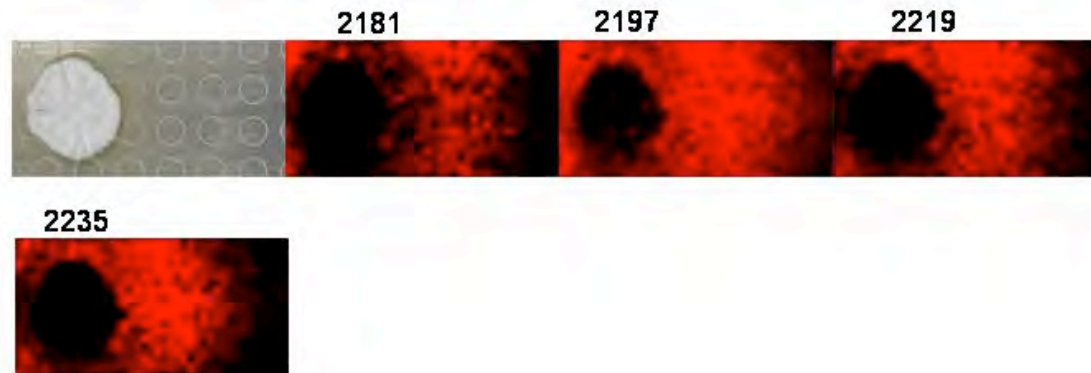
IMS 19. *Streptomyces albus* grown on ISP-2 media.

***Streptomyces ghanaensis* 14672 on ISP-2**



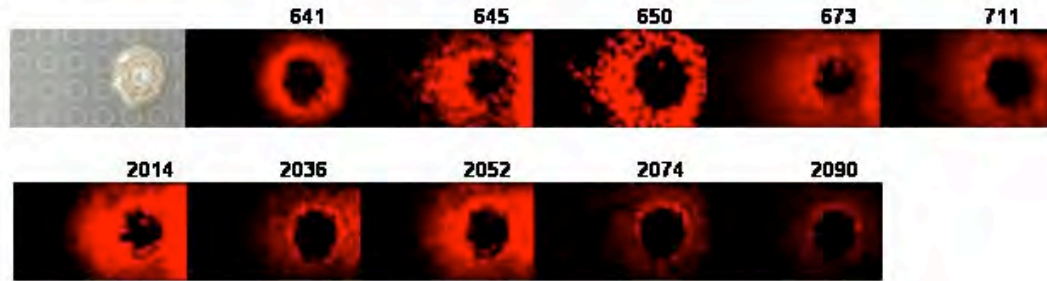
IMS 20. *Streptomyces ghanaensis* grown on ISP-2 media.

*S. clavuligerus* ATCC 53653 on ISP2



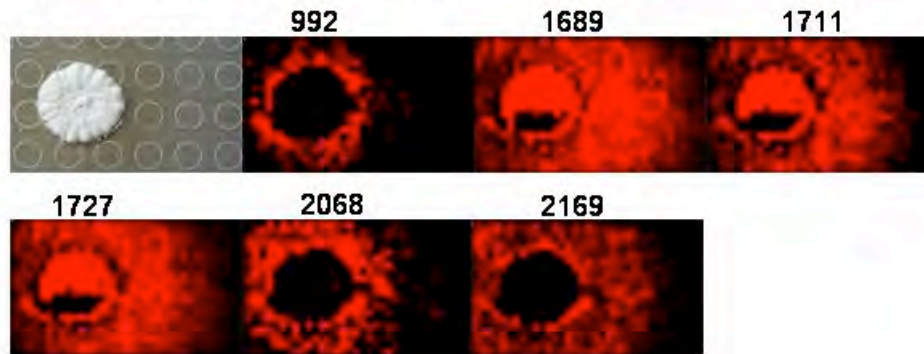
IMS 21. *Streptomyces clavuligerus* grown on ISP-2 media.

***Streptomyces pristinispiralis* ATCC25486 on ISP-2**



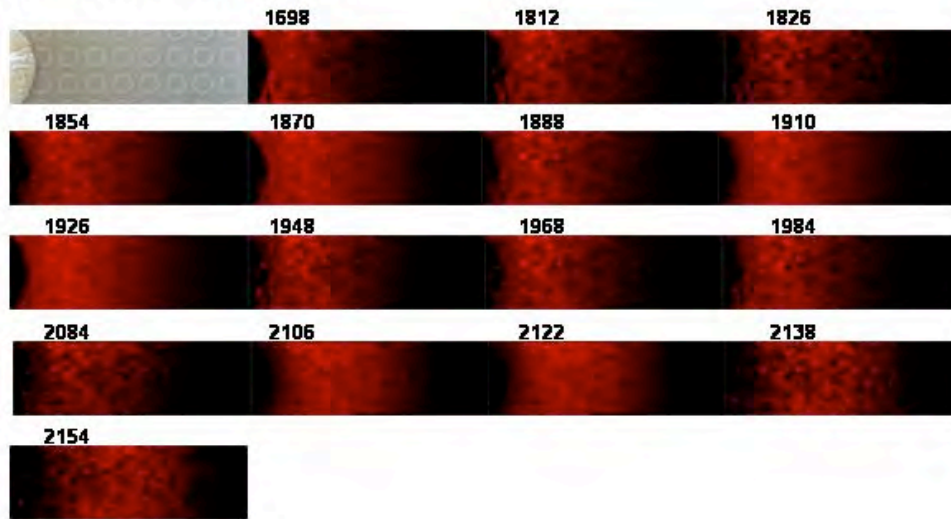
IMS 22. *Streptomyces pristinispiralis* grown on ISP-2 media.

*Streptomyces viridochromogenes* 40736 on ISP-2



IMS 23. *Streptomyces viridochromogenes* grown on ISP-2 media.

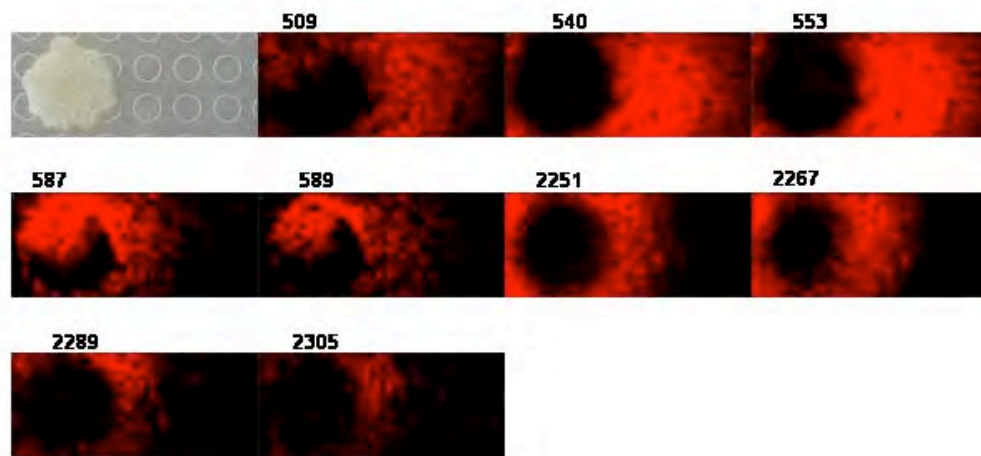
***Streptomyces sviveus* ATCC29083**



IMS 24. *Streptomyces sviveus* grown on ISP-2 media.

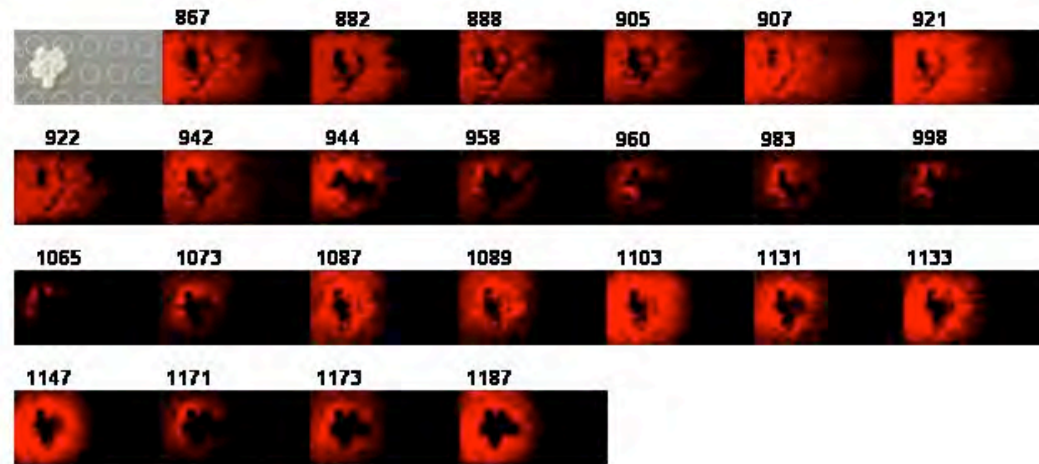


***Streptomyces* sp. SPB78**



IMS 25. *Streptomyces* sp. SPB78 grown on ISP-2 media.

***Kutzneria* sp. 744 on ISP-2**



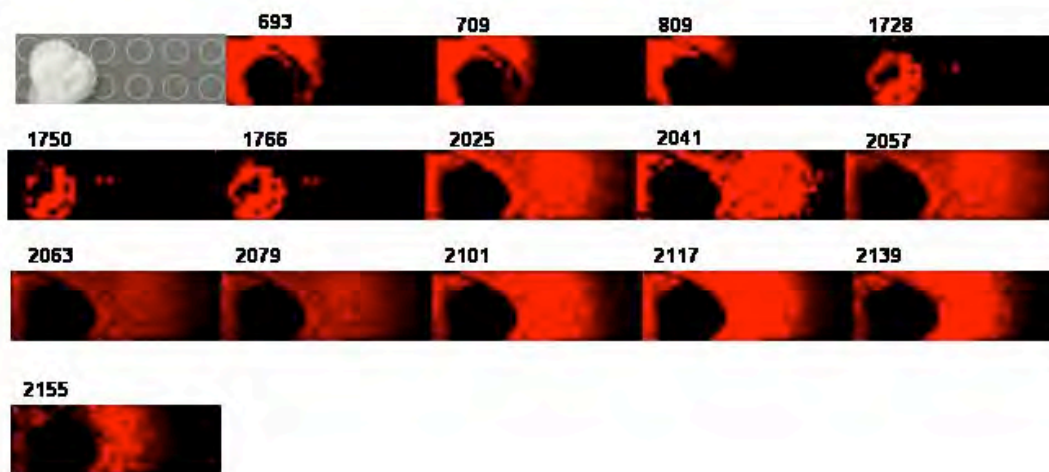
IMS 26. *Kutzneria* sp. 744 grown on ISP-2 media.

***Actinoplanes teichomyceticus* PSK0532 on ISP-2**



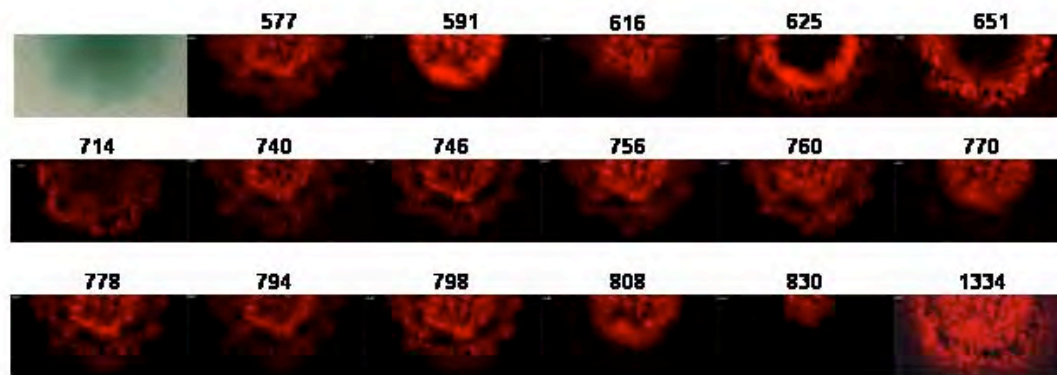
IMS 27. *Actinoplanes teichomyceticus* grown on ISP-2 media.

***Streptovercillium griseovercillatum* ATCC 31499 on ISP-2**



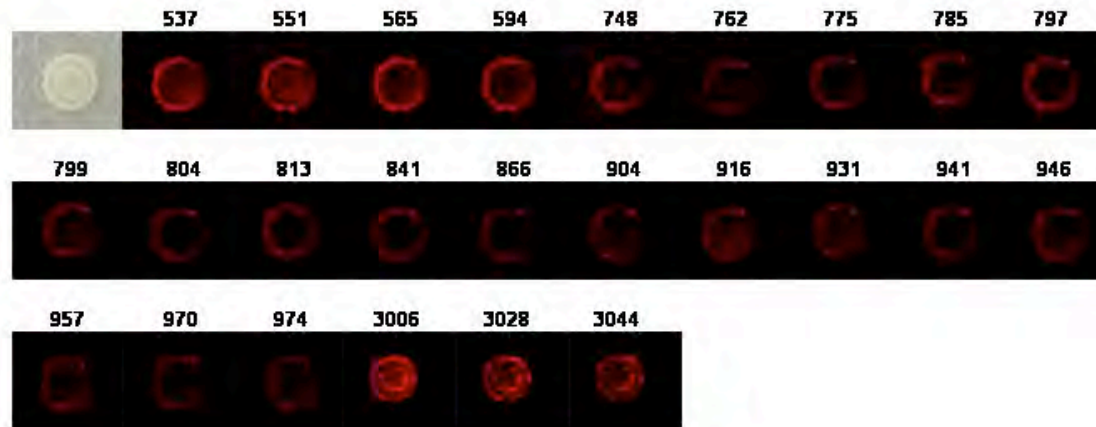
IMS 28. *Streptovercillium griseovercillatum* on ISP-2 media. The ion at 2041 Da was identified to be the [M+H] of Cinnamycin by tandem mass spectrometry (Supplemental Figure 2).

***Pseudomonas aeruginosa* PAO on ISP-2**



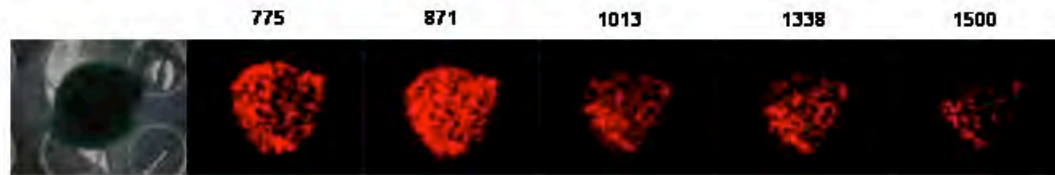
IMS 29. *Pseudomonas aeruginosa* grown on ISP-2 media.

***Staphylococcus aureus* USA300 on ISP-2**



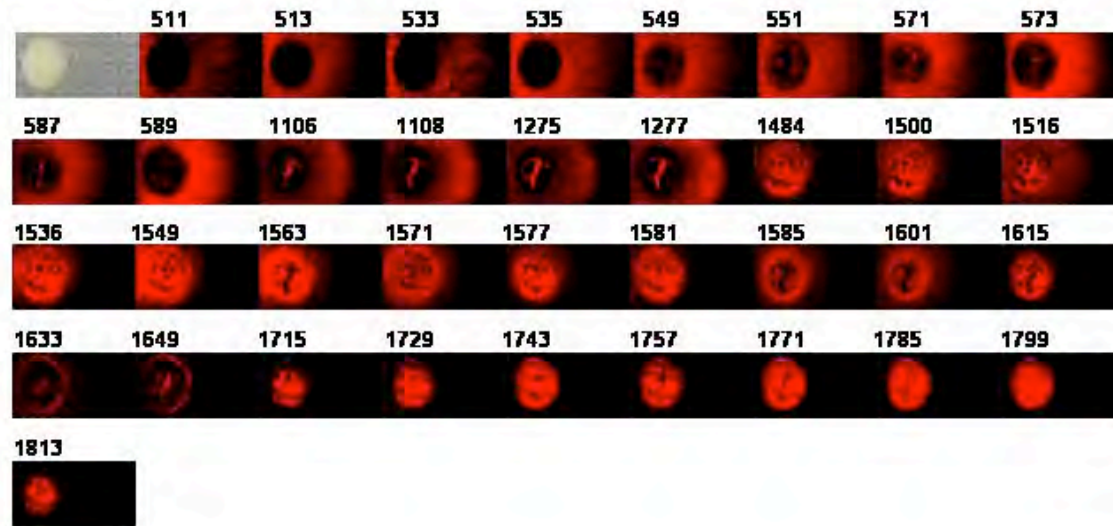
IMS 30. *Staphylococcus aureus* USA300 grown on ISP-2 media. The ion at 3006 was indentified to be  $\delta$ -toxin (PSM $\lambda$ ) by tandem mass spectrometry.

**Nostoc sp. PCC 7120 on BG-11**



IMS 31. Nostoc sp. grown on BG-11 media.

***Lysobacter enzymogenes* C3 on ISP-2**



IMS 32. *Lysobacter enzymogenes* grown on ISP-2 media.

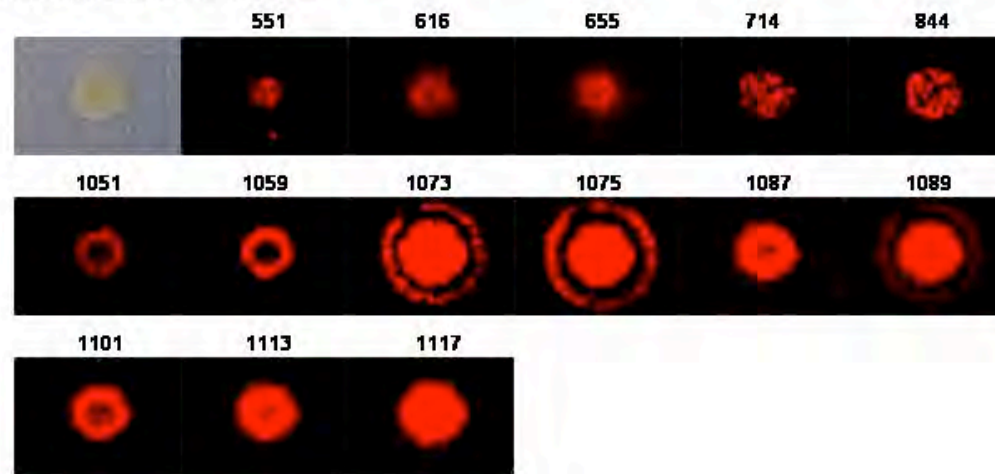


***Bacillus subtilis* 3610 on ISP-2**



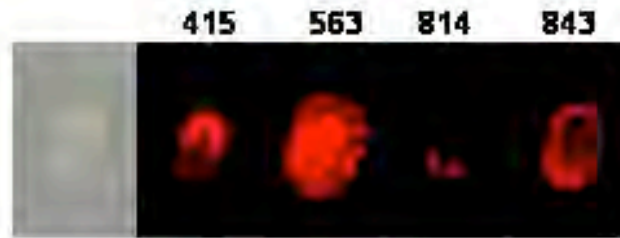
IMS 33. *Bacillus subtilis* grown on ISP-2 media. The ion at 1075 Da was identified to be the [M+K] of Surfactin by tandem mass spectrometry. IMS did not detect the [M+H] (Supplemental Figure 2). The ion at 1545 Da was identified to be the [M+K] of Plipastatin by tandem mass spectrometry. IMS did not detect the [M+H] (Supplemental Figure 2).

***Bacillus pumilus* GNJ762 on ISP-2**



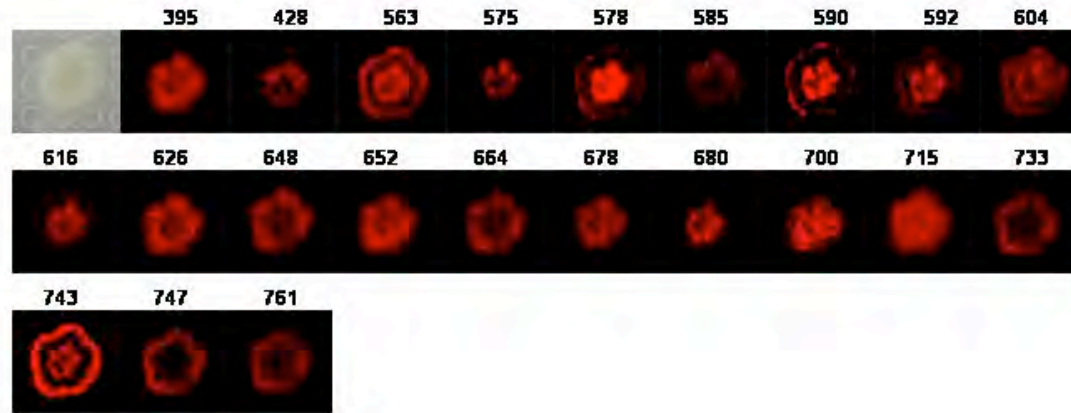
IMS 34. *Bacillus pumilus* grown on ISP-2 media. The ion at 1075 Da was identified to be the [M+K] of Surfactin by tandem mass spectrometry. IMS did not detect the [M+H] (Supplemental Figure 2).

***Rhizobium leguminosarum* S36 on ISP-2**



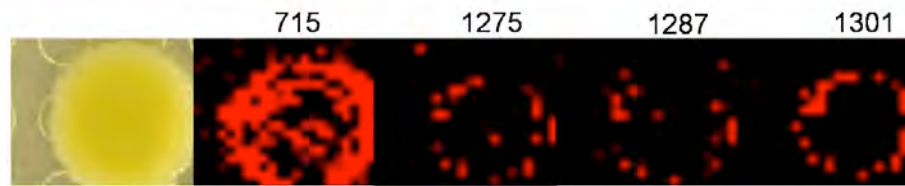
IMS 35. *Rhizobium leguminosarum* grown on ISP-2 media.

***Pseudomonas fluorescens* PFO-1 on ISP-2**



IMS 36. *Pseudomonas fluorescens* grown on ISP-2 media.

***Mycobacterium smegmatis* MC2 155**



IMS 37. *Mycobacterium smegmatis* grown on ISP-2 media.

**Supplementary Table 2 Characterization of named ions in this study.**

	<b>Streptorubin B<sub>2</sub><sup>+</sup></b>	<b>error ppm</b>	<b>Surfactin-C<sub>15</sub><sup>+</sup></b>	<b>error ppm</b>	<b>CDA<sub>2</sub><sup>+</sup></b>	<b>error ppm</b>	<b>Plipastatin-C<sub>17</sub>-Val<sup>+</sup></b>	<b>error ppm</b>	<b>SapB<sub>2</sub><sup>+</sup></b>	<b>error ppm</b>	<b>Beauvericin</b>	<b>error ppm</b>
<b>Molecular formula</b>	C <sub>25</sub> H <sub>33</sub> N <sub>3</sub> O		C <sub>53</sub> H <sub>93</sub> N <sub>7</sub> O <sub>13</sub>		C <sub>67</sub> H <sub>78</sub> N <sub>14</sub> O <sub>26</sub>		C <sub>75</sub> H <sub>116</sub> N <sub>12</sub> O <sub>20</sub>		C <sub>84</sub> H <sub>139</sub> N <sub>25</sub> O <sub>29</sub> S <sub>2</sub>		C <sub>45</sub> H <sub>57</sub> N <sub>3</sub> O <sub>9</sub>	
<b>Calcd. [M+H]<sup>+</sup></b>	392.2702		1036.6910		1497.5290		1505.8507		2026.9690		784.4173	
<b>Calcd. [M+K]<sup>+</sup></b>	430.2261		1074.6468		1536.4849		1543.8066		2064.9249		822.3732	
<b>Calcd. [M+Na]<sup>+</sup></b>	414.2521		1058.6729		1519.5109		1527.8327		2048.9510		806.3993	
<b>Found w/ MALDI [M+H]<sup>+</sup></b>	392.2720	4.6	ND		1497.5029	17.5	ND		2026.9706	0.8	ND	
<b>Found w/ MALDI [M+K]<sup>+</sup></b>			1074.6710	22.5	1536.4749	6.5	1543.8571	32.7	2064.9165	4.1		
<b>Found w/ MALDI [M+Na]<sup>+</sup></b>			1058.6765	3.4	1519.4967	9.4	1527.8605	18.2				
<b>Found w/ FTMS [M+H]<sup>+</sup></b>	392.2698	1.0			1497.5316	1.7	1505.8563	3.7			784.4203	3.8
<b>Found w/ FTMS [M+K]<sup>+</sup></b>			1074.6469	0.1							822.3754	2.7
<b>Found w/ FTMS [M+Na]<sup>+</sup></b>											806.4056	7.8
<b>MS2</b>	TOFTOF		TOFTOF		TOFTOF		TOFTOF		TOFTOF		ITMS2	
<b>Representative in imaging</b>	392		1075		1536		1545		2027		806	

Supplementary Table 2. Characterization of named ions in this study (continued)

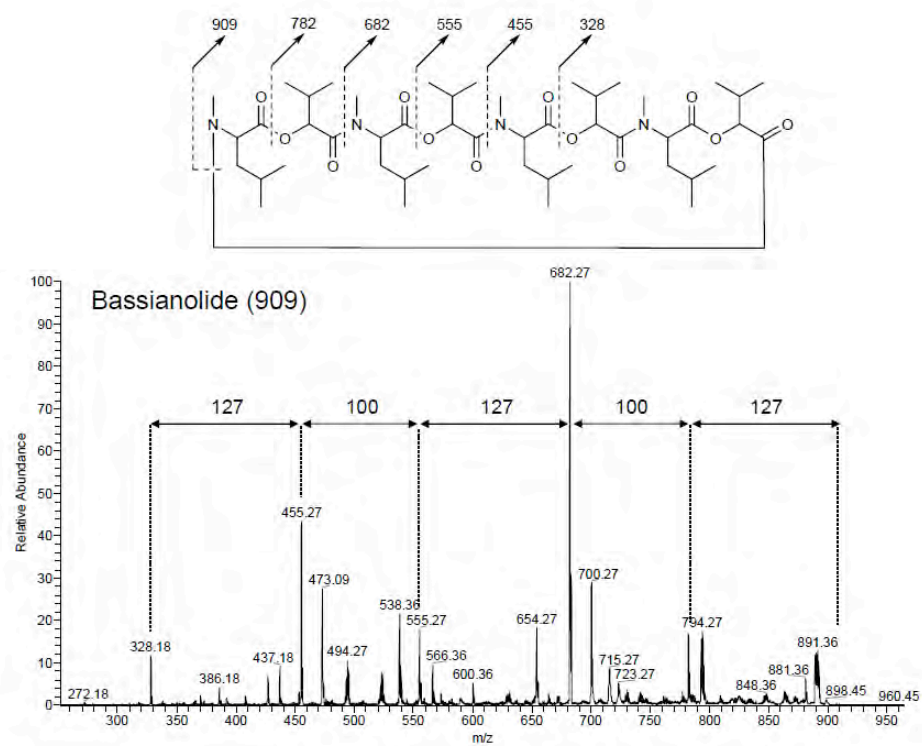
	Bassianolide	error ppm	Etamycin	error ppm	Cinnamycin	error ppm	Maltophytin	error ppm	Dihydromaltophytin	error ppm	Pheophytin A	error ppm
<b>Molecular formula</b>	C <sub>48</sub> H <sub>84</sub> N <sub>4</sub> O <sub>12</sub>		C <sub>44</sub> H <sub>62</sub> N <sub>8</sub> O <sub>11</sub>		C <sub>89</sub> H <sub>125</sub> N <sub>25</sub> O <sub>25</sub> S <sub>3</sub>		C <sub>29</sub> H <sub>38</sub> N <sub>2</sub> O <sub>6</sub>		C <sub>29</sub> H <sub>40</sub> N <sub>2</sub> O <sub>6</sub>		C <sub>55</sub> H <sub>74</sub> N <sub>4</sub> O <sub>5</sub>	
<b>Calcd. [M+H]<sup>+</sup></b>	909.6164		879.4616		2040.8519		511.2808		513.2965		871.5737	
<b>Calcd. [M+K]<sup>+</sup></b>	947.5723		917.4175		2078.8078		549.2367		551.2523		909.5296	
<b>Calcd. [M+Na]<sup>+</sup></b>	931.5983		901.4436		2062.8338		533.2628		535.2784		893.5557	
<b>Found w/ MALDI [M+H]<sup>+</sup></b>											871.5559	20.4
<b>Found w/ MALDI [M+K]<sup>+</sup></b>												
<b>Found w/ MALDI [M+Na]<sup>+</sup></b>												
<b>Found w/ FTMS [M+H]<sup>+</sup></b>	909.6212	5.3	879.4642	3.0	2040.8566	2.3	511.2819	2.2	513.2971	1.2		
<b>Found w/ FTMS [M+K]<sup>+</sup></b>	947.5770	5.0	917.4245	7.6	2078.8156	3.8						
<b>Found w/ FTMS [M+Na]<sup>+</sup></b>	931.6022	4.2	901.4495	6.5			533.2651	4.3	535.2804	3.7		
<b>MS2</b>	ITMS2		ITMS2		ITMS2		ITMS2		ITMS2		TOFTOF	
<b>Representative in imaging</b>	932		879		2041		511		513		871	

\* [Described](#) in Yang Y.L., Xu Y., Straight P., Dorrestein P.C. Translating metabolic exchange with imaging mass spectrometry. *Nature Chemical Biology*. 5, 885-887. 2009



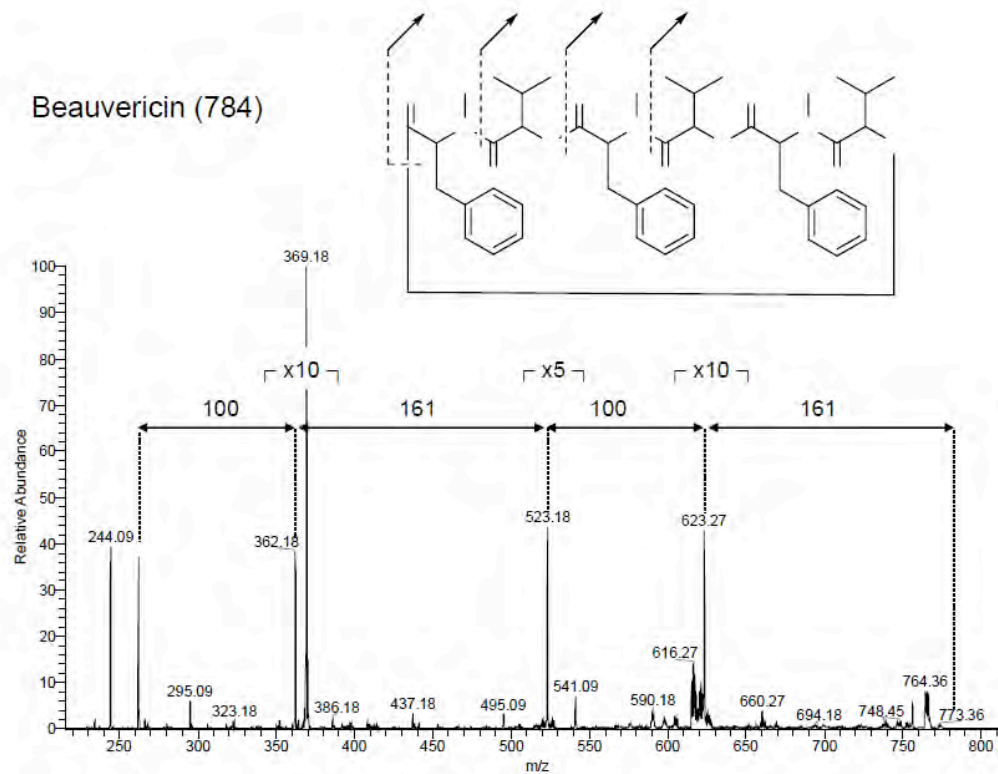


**Supplementary Figure 2. The annotated known compounds observed by TLA-IMS**

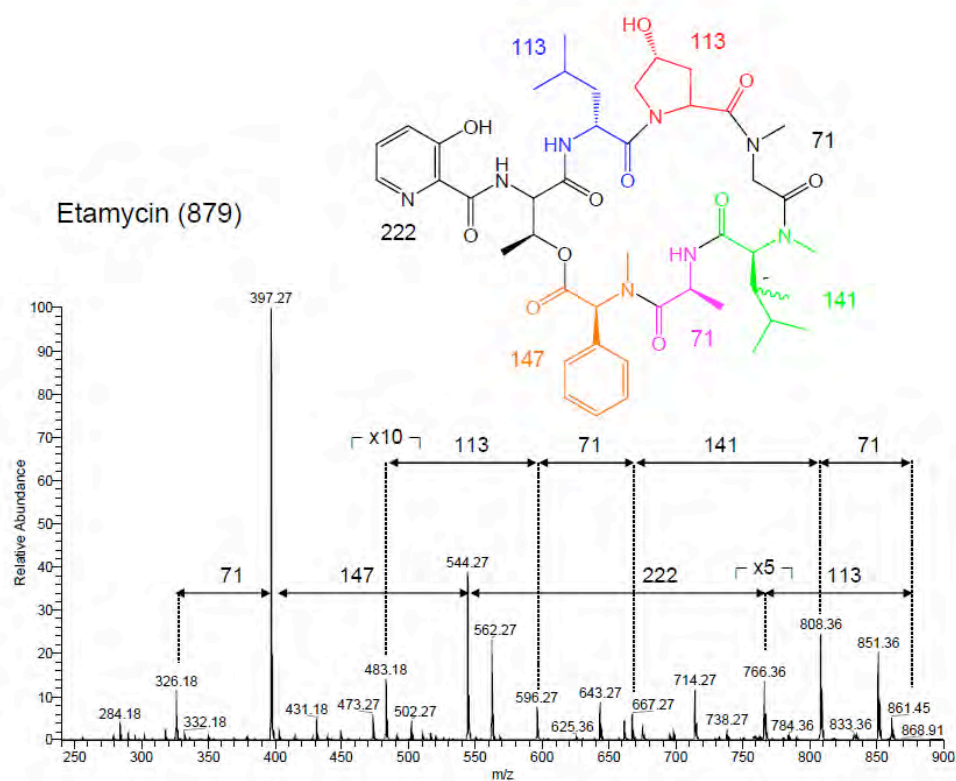


Spectrum 1. Bassianolide. FTICR-MS was used to identify the  $[M+H]^+$  ion at 5.3 ppm mass error. Thereafter, tandem mass spectrometry using a LTQ-MS was performed to identify fragment ions. The number in brackets within the figures is the molecular mass of the compound.

Beauvericin (784)

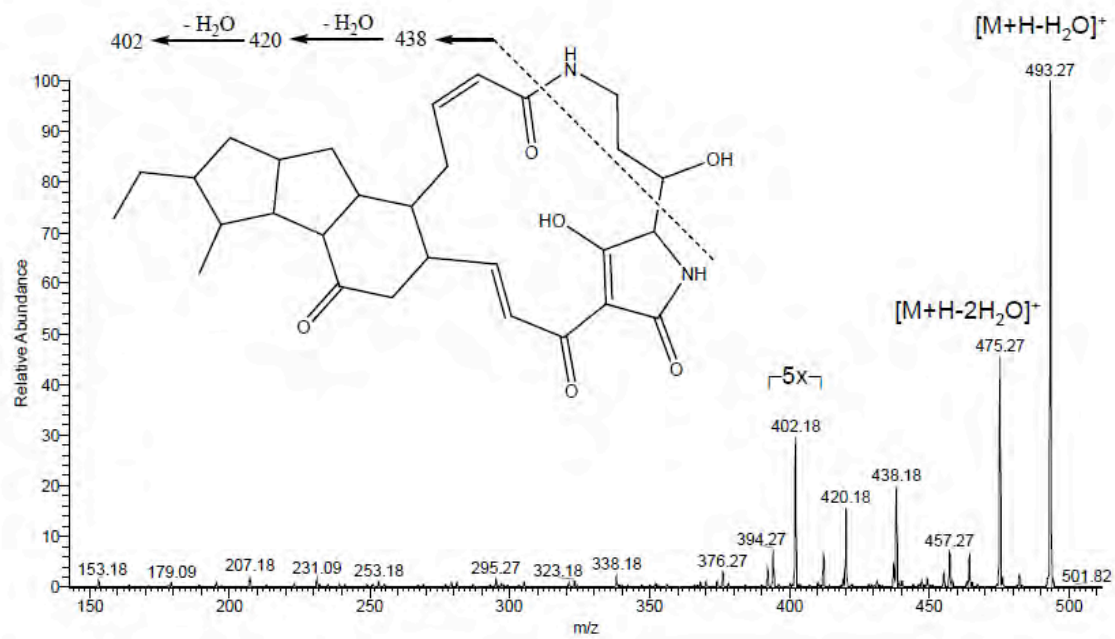


Spectrum 2. Beauvericin. FTICR-MS was used to identify the [M+H] ion at 3.8 ppm mass error. Thereafter, tandem mass spectrometry using a LTQ-MS was performed to identify fragment ions. The number in brackets within the figures is the molecular mass of the compound.



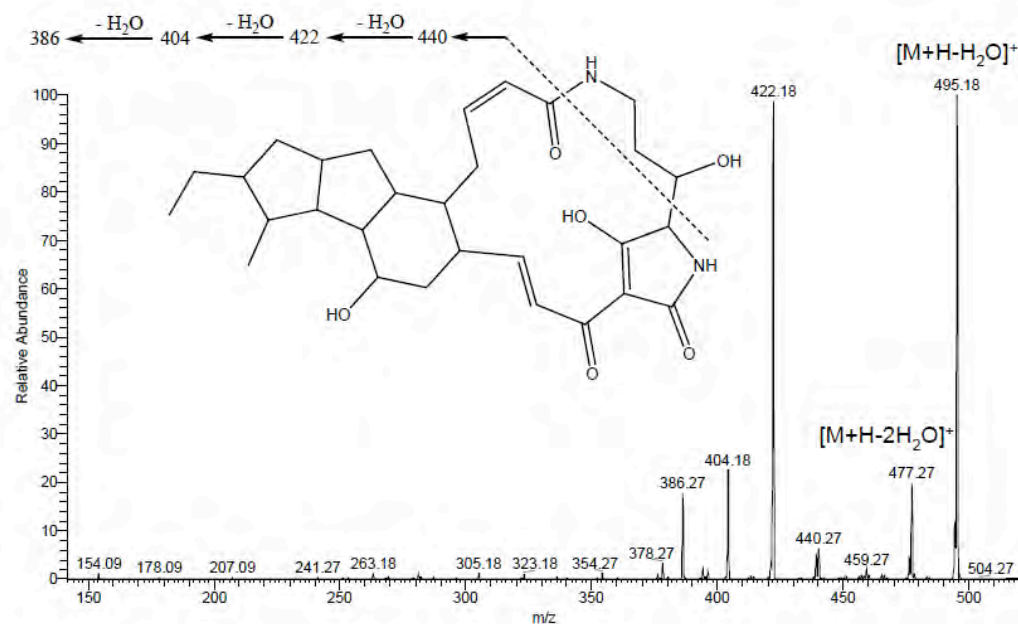
Spectrum 3. Etamycin. FTICR-MS was used to identify the  $[M+H]^+$  ion at 3 ppm mass error. Thereafter, tandem mass spectrometry using a LTQ-MS was performed to identify fragment ions. Identified fragments are depicted in the structure insert. The number in brackets within the figures is the molecular mass of the compound.

### Maltophytin (511)

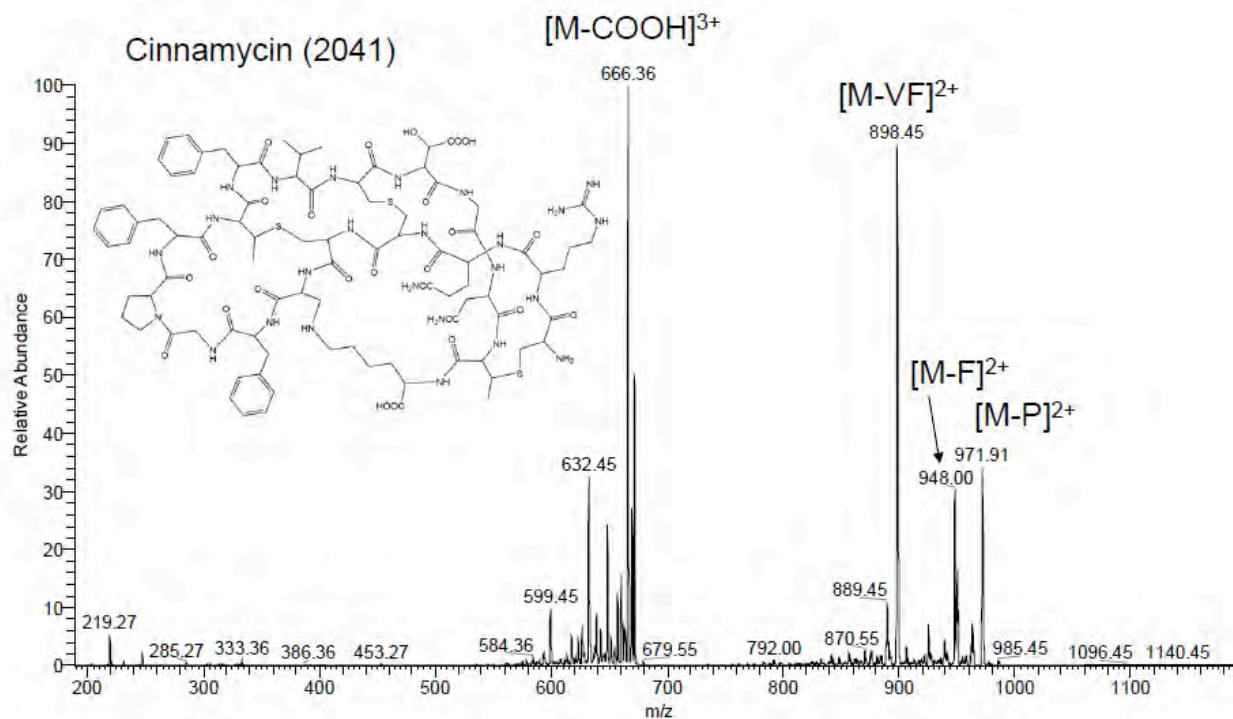


Spectrum 4. Maltophytin. FTICR-MS was used to identify the  $[M+H]$  ion at 2.2 ppm mass error. Thereafter, tandem mass spectrometry using a LTQ-MS was performed to identify fragment ions. Structure insert depicts identified fragment ions. The number in brackets within the figures is the molecular mass of the compound.

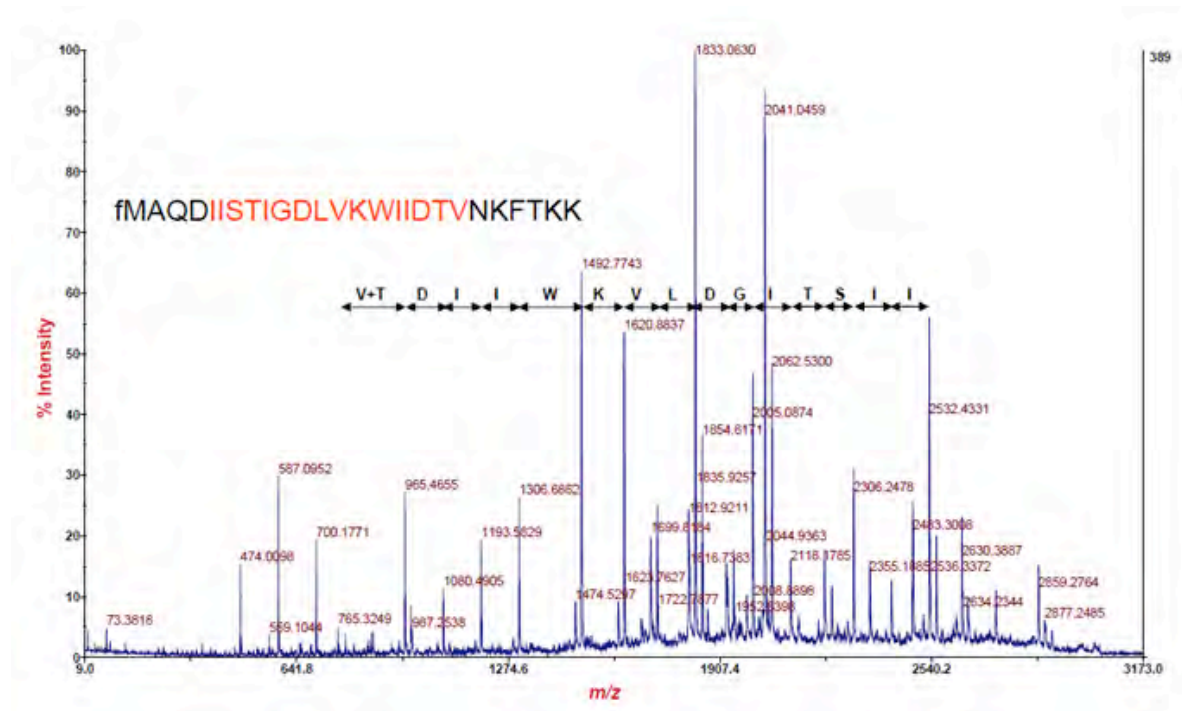
### Dihydromaltophytin (513)



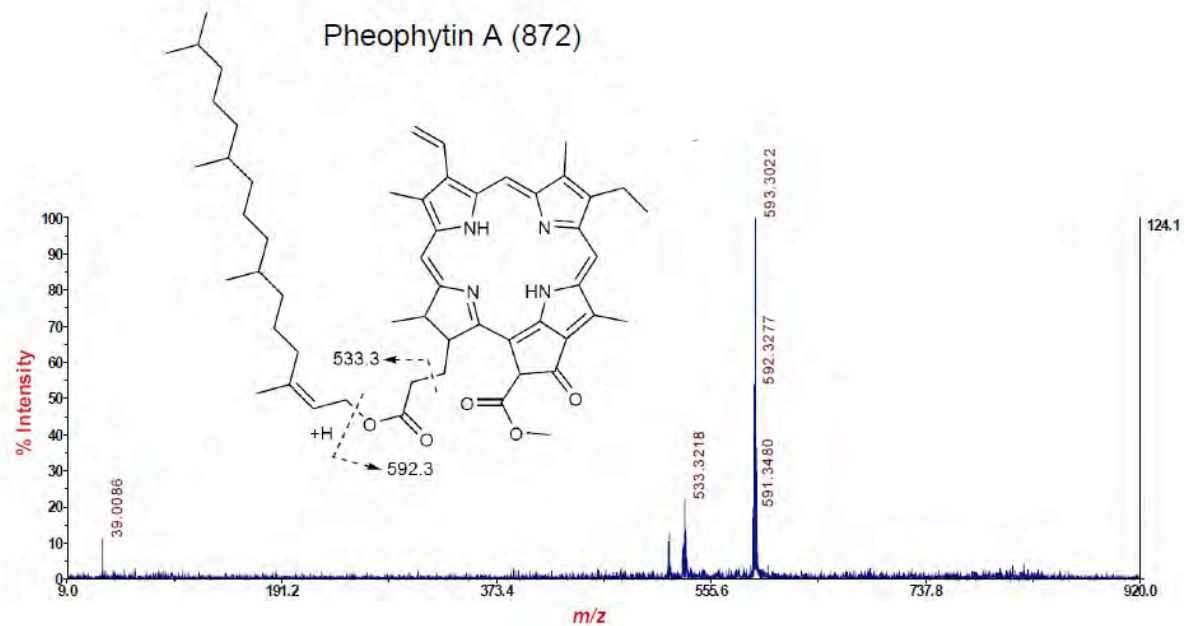
Spectrum 5. Dihydromaltophytin. FTICR-MS was used to identify the  $[M+H]$  ion at 1.2 ppm mass error. Thereafter, tandem mass spectrometry using a LTQ-MS was performed to identify fragment ions. Structure insert depicts identified fragment ions. The number in brackets within the figures is the molecular mass of the compound.



Spectrum 6. Cinnamycin. FTICR-MS was used to identify the ion corresponding to Cinnamycin at 2.3 ppm mass error. Thereafter, tandem mass spectrometry using a LTQ-MS was performed to identify fragment ions as indicated on the spectrum. The number in brackets within the figures is the molecular mass of the compound.

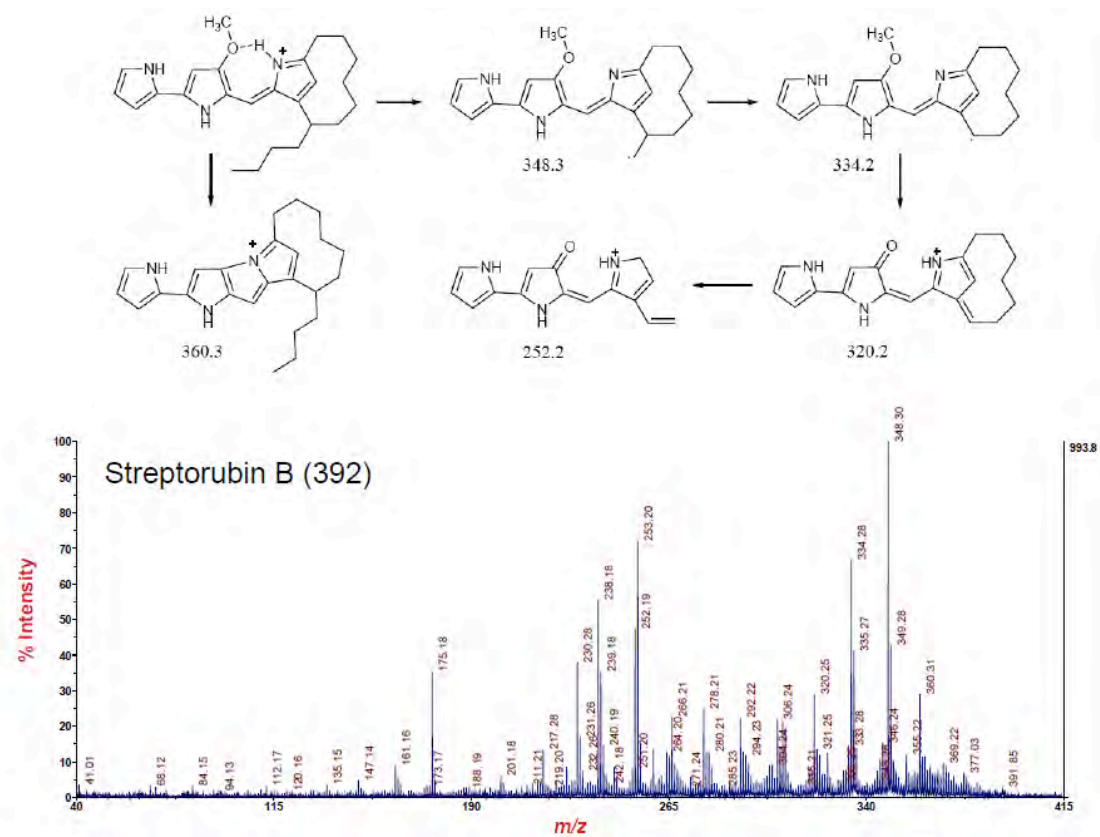


Spectrum 7.  $\delta$ -toxin. The ABI 4800 MALDI-TOF/TOF was used to identify the  $[M+H]^+$  ion at 45.6 ppm mass error. Thereafter, tandem mass spectrometry was performed to fragment the target ion. A sequence tag was generated as indicated within the spectrum and BLAST analysis was used to identify the ion as  $\delta$ -toxin.

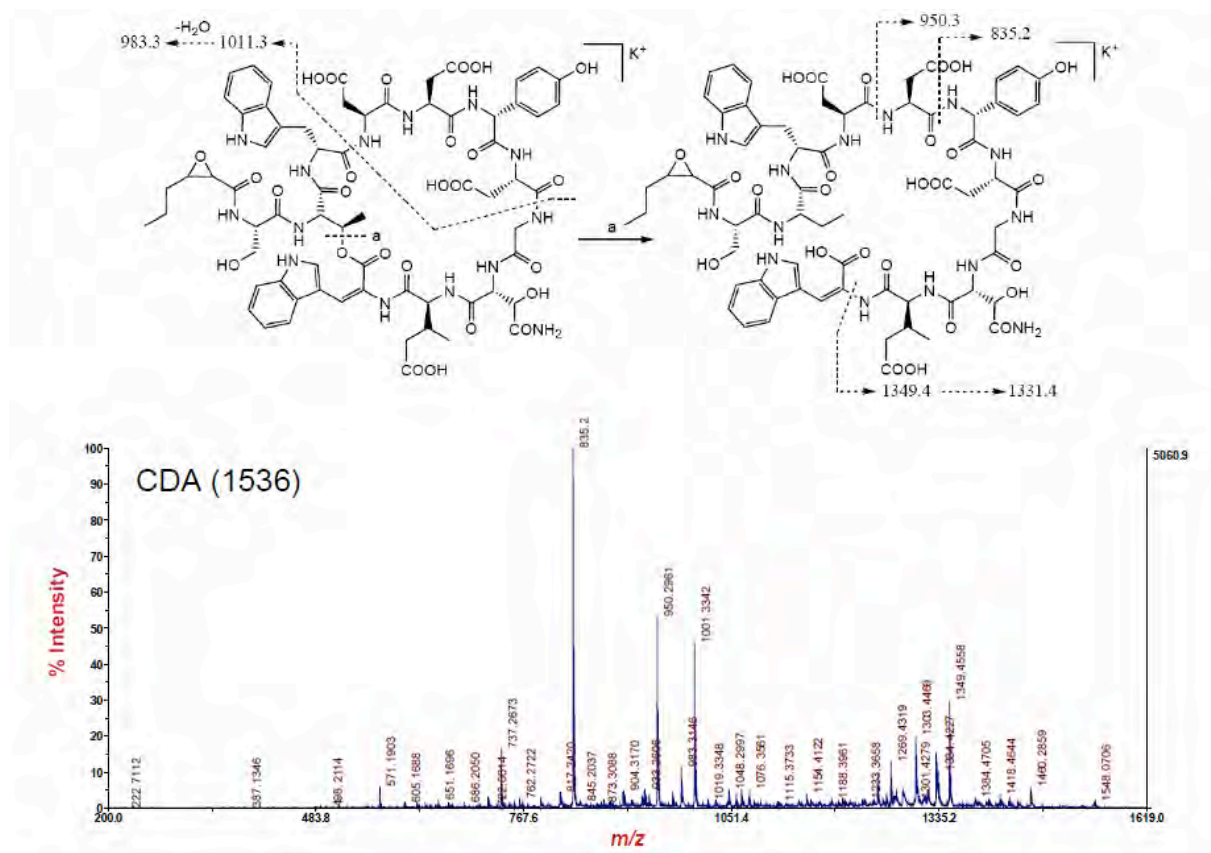


Spectrum 8. Pheophytin A. The ABI 4800 MALDI-TOF/TOF was used to identify the  $[M+H]$  ion at 20.4 ppm mass error. Thereafter, tandem mass spectrometry was performed to fragment the target ion. Ion fragments were identified and are indicated on the structure insert. The number in brackets within the figures is the molecular mass of the compound.





Spectrum 9. Streptorubin B. The ABI 4800 MALDI-TOF/TOF was used to identify the [M+H] ion at 4.6 ppm mass error and further verified with FTICR-MS at 1.0 ppm mass error. Thereafter, tandem mass spectrometry was performed to fragment the target ion. Ion fragments were identified and are indicated on the structure insert. The number in brackets within the figures is the molecular mass of the compound.



Spectrum 10. CDA. The ABI 4800 MALDI-TOF/TOF was used to identify the  $[M+H]^+$  ion at 17.5 ppm mass error and further verified with FTICR-MS at 1.7 ppm mass error. Thereafter, tandem mass spectrometry was performed to fragment the target ion. Ion fragments were identified and are indicated on the structure insert. The number in brackets within the figures is the molecular mass of the compound.